



## PHYTOEXTRACTION OF HEAVY METALS BY *Ricinus communis* IN SOIL AMENDED WITH CHELANTS AND POULTRY MANURE



Raymond A. Wuana<sup>1</sup>, Lami A. Nnamonu<sup>1</sup>, Adams U. Itodo<sup>1</sup> and Gabriel T. Buluku<sup>2</sup>

<sup>1</sup>Department of Chemistry, University of Agriculture Makurdi, Benue State, Nigeria

<sup>2</sup>Department of Science Laboratory Technology (SLT), Gboko Polytechnic, Benue State, Nigeria

\*Corresponding author: [raynewton@yahoo.com](mailto:raynewton@yahoo.com)

Received: April 15, 2019 Accepted: August 17, 2019

**Abstract:** Pot experiments were designed to investigate the response of *R. communis* to heavy metals phytoextraction tested under single or mixed chelants and poultry manure assisted scenarios. This was achieved when a moderately contaminated sandy loamy soil was stressed with a 500 mgkg<sup>-1</sup> conjointly metal (Cd, Cu, Pb), blended with; single or mixed chelant (EDTA and oxalic acid) and poultry manure. After growth, the maximum heights and leaf breadths of *R. communis* were; 5.01-69.0 and 1.90-38.4 cm, with plants typically luxuriant and greenish. pH, bulk density, electric conductivity, organic carbon, of the parent soil were determined. The results revealed that pH (6.21), (9.30 cmol Kg<sup>-1</sup>), BD (1210 kg/m<sup>3</sup>), OM (8.63 %) were found in the parent soil used. Pseudo-total metal concentration (mgkg<sup>-1</sup>) of the metals (Cd, Cu and Pb) were; 10.2, 8.93 and 5.22, respectively. Maximum tissue metal concentrations in the various potting media were: soil-metal-PM-EDTA+OX (436.0-490.6 mg Cd/kg; 453.2-475.5 mg Cu/kg; 452.3-469.8 mg Pb/kg. Maximum values for Cd, Cu and Pb root bioaccumulation factor (RB<sub>r</sub>), shoot bioaccumulation factor (SB<sub>r</sub>), and translocation factor (T<sub>r</sub>) were; 0.68, 0.29 and 0.42; 0.67, 0.28 and 0.42; and 0.66, 0.27 and 0.41, respectively. Pot experiments revealed high metal transferabilities with no apparent phytotoxic symptoms in *R. communis* at the doses applied, suggesting some degree of tolerance to the metals. Overall, the binary chelant treatments were less toxic for *R. communis* growth and enhanced metal accumulation in shoots to a greater extent than the single chelant scenarios, but more so when EDTA was present in the binary combination.

**Keywords:** Heavy metals, *Ricinus communis*, phytoextraction, poultry manure, chelants

### Introduction

The pollution of soil is a crucial matter that has attracted considerable public attention over the past few decades (Ahmadpour *et al.*, 2012). This is because pollution is a threat for the survival of mankind and the most important dispute of our era (Wang *et al.*, 2004). Geological and anthropogenic activities are sources of heavy metal contamination. Sources of anthropogenic metal contamination include industrial effluents, fuel production, mining, smelting processes, military operations, small-scale industries (including battery production, metal products, metal smelting and cable coating industries), brick kilns and coal combustion (Dembitsky, 2003). Heavy metals that are most commonly found in contaminated soils (but not necessarily listed in order of abundance) are arsenic (As), cadmium (Cd), copper (Cu), chromium (Cr), mercury (Hg), lead (Pb), nickel (Ni), and zinc (Zn) (Wuana and Okieimen, 2011).

The mobilization of heavy metals by man through extraction from ores, and also the disposal of municipal sewage and agricultural practices have led to the release of these elements into the environment.

Phytoremediation is frequently listed among the best demonstrated available environmentally less-invasive in situ technologies regarded as primary remedies for restoring the quality and functionality of soils disturbed by toxic heavy metals (Pedron and Petruzzelli, 2011; Wuana and Mbasugh, 2013). Phytoextraction of metals offers benefits such as; decontamination of land, exploitation of more widely-dispersed resources of surface metals and recovery of metals from ore bodies with low metal concentration. Phytoextraction has emerged as a promising, cost-effective alternative to the conventional engineering-based remediation methods. It is a relatively recent technology and is perceived efficient, novel, eco-friendly, and solar-driven technology with good public acceptance. It is a method which, plants are used to remove metals from soils, transport and concentrate them in above-ground biomass (Padmavathamma and Loretta 2007). Phytoremediation is an area of active current research. New efficient metal hyperaccumulators are being explored for

applications in phytoremediation and phytomining. Plants to be used for phytoextraction should have: (a) tolerance to high concentrations metals, (b) high metal-accumulation capability, (c) heavy biomass, (d) ability to grow fast and a (e) profuse root system (f) Tolerance to the toxic effects of the target heavy metals. (g) Resistance to pathogens and pests. (h) Repulsion to herbivores to avoid food chain contamination. The success of phytoextraction depends especially on the plant's ability (a) to accumulate biomass rapidly, and (b) to store large quantities of the up taken metals in the shoot tissue (McGrath, 1998; Blaylock and Huang, 2000)

Chelate-enhanced, phytoextraction is based on the fact that the application of organic and inorganic chelating agents to the soil to significantly enhance metal accumulation by plants (Garbisu and Alkorta, 2001). The formation of chelates prevents precipitation and sorption of the metals, thereby maintaining their availability for plant uptake (Salt *et al.*, 1995). Chelants possess varying chemical affinities for different metals, and so the presence of metal mixtures with their synergistic or antagonistic interactions may impair the beneficial effects of the chelants (do Nascimento *et al.*, 2006). The solubilization of heavy metals through inorganic agents relies mainly on desorption (Bru'mmer *et al.*, 1986). Heavy metal solubility in soils is mainly controlled by the soil reaction (pH), the amount and kind of sorption sites, and the total amount of heavy metals in the soil (Hornburg and Bru'mmer, 1993; Gray *et al.*, 1999).

Castor (*Ricinus communis* L.) is one of the oldest cultivated oil crops in the world (Ruwanthi, 2012). The castor oil plant (*Ricinus communis*) is a species of flowering plant in the spurge family, Euphorbiaceae. Castor bean (*Ricinus communis* L.) has been extensively exploited for their heavy metal tolerance and phytoremediation potential (Adhikari and Kumar, 2012). Recent reports have indicated that *R. communis* can be a multi-tasking for use in phytoremediation of soils contaminated by toxic heavy metals and carbon abatement technology due to its relatively high growth rate, profuse root system, prolific biomass yield, metal tolerance and metal accumulation and high carbon fixation (Rajkumar

and Freitas, 2008; Shi and Cai, 2009; Huang *et al.*, 2011; Miniño *et al.*, 2014; Zhang *et al.*, 2014; Wuana *et al.*, 2016). The present study was aimed at assessing the growth potential and response of *Ricinus communis* under the influence of poultry manure and chelants (single and binary) in soils with Cd, Cu and Pb.

### Materials and Methods

Chemicals and apparatus used for the study included cadmium nitrate, coppers sulphate, lead nitrate, oxalic acid and disodium salt of ethylenediaminetetraacetic acid. Allorganic chelants used were of Sigma-Aldrich patent. Atomic absorption spectrophotometer (Buck Scientific Model 2006A, Norwalk, Connecticut, US).

### Description of study area

This study was carried out on a parent soil sample collected from the extended Judges' quarters layout sited in Makurdi, north-central of Nigeria. It is located at 7° 48' N and longitude 8° 37' E in the lower Benue River Basin, Annual rainfall is 1200–1650 mm distributed between March/April and October/November, followed by a marked dry season (of up to 4 months). The ranges of daily maximum and minimum temperatures are 30–34°C and 22–24°C, respectively, during the rainy season and 33–37°C and 18–24°C, respectively, in the dry season. Daily global irradiation and mean hours of insulation are, respectively, 314–433 cal cm<sup>2</sup> day<sup>-1</sup> and 4.00–7.74 h (Sha' Ato *et al.*, 2002).

### Soil sampling, characterization, artificial contamination and treatment with chelant and/or manure

In this study five surface (0–20 cm) soil samples were randomly collected using a chrome-plated trowel. The soil samples were air dried, ground, sieved to <2-mm particle size, composited, and stored in polythene bags as the parent soil (PS). A composite sample of poultry manure (PM) collected from a poultry farm in Makurdi and was kept for two weeks to age. The physicochemical properties of the parent soil and the raw poultry manure used as an amendment were determined as described by Wuana *et al.* (2012). Pseudototal Cd, Cu, and Pb contents were determined by digestion with aqua regia (HCl–HNO<sub>3</sub>) which was subsequently followed by metal assay using AAS.

In order to study the effect(s) of metal dose on *R. communis* growth, the parent soil was spiked with a solution (500 mg/kg) conjointly dose of Cd, Cu, and Pb and a poultry manure (15% w/w). A solution of 100 mL increasing doses (mmol chelant/kg soil): 0, 5, 10, 15 of disodium salt of EDTA and/or sodium oxalate solutions was added over a thin layer the 5.0-kg portions of the metal-manure-spiked soil, to furnish a soil-metal-poultry manure-EDTA, soil-metal-poultry manure-sodium oxalate and, soil-metal-poultry manure-EDTA and sodium oxalate potting media. Each treatment was performed in triplicate and incubated for 2 weeks at ambient temperature to simulate field conditions. Sub-samples of the amended soils were digested and pseudototal Cd, Cu, and Pb contents determined by AAS analysis.

### Pot experiments with *R. communis*

Three seeds of *R. communis*, previously cold treated (10°C) for 12 h according to Wuana *et al.* (2013) to break dormancy, synchronize germination prior to pot experiments (Revathi *et al.*, 2010). Pots were placed in a completely randomized design. Five seeds were sown in each pot (volume = 5000 mL) of 5.0 kg soil and a week after germination, the seedlings were thinned to three. Surface irrigation with deionized water was employed to water the plants during growth and no fertilizers were applied. Night and day sequence was naturally obtained by maintaining the pots in an open area. The plants were monitored for twelve (12) weeks for changes in appearance (colour), height, and leaf breadth. Prior to harvest, plants were left without watering for one (1) day. They plants

were carefully uprooted from the soils after the 90<sup>th</sup> day, separated into roots and shoots, rinsed with deionised water, and dried at 110°C for 72 h. The concentrations of Cd, Cu and Pb (mg kg<sup>-1</sup>dw of *R. communis* biomass) was determined by HNO<sub>3</sub>–H<sub>2</sub>O<sub>2</sub> digestion (Nolan *et al.*, 2005) followed by atomic absorption spectrophotometric measurements.

### Plant biomass digestion and heavy assay in *Ricinus communis*

A one (1) gram dried and ground *R. communis* biomass sample was digested with a mixture of 4.0 mL of 65% v/v HNO<sub>3</sub> and 2.0 mL of 35% v/v H<sub>2</sub>O<sub>2</sub>. After evaporation, 4.0 mL of concentrated HNO<sub>3</sub> and 2.0 mL of concentrated H<sub>2</sub>O<sub>2</sub> were added to the residue and heated until a clear digest appeared. The digestion time of 3 h at 130°C was employed. The digest was made up to the 5.0 mL with 1.0 M HNO<sub>3</sub> (Wuana *et al.*, 2013). The concentrations for Cd, Cu, and Pb were then estimated using Atomic Absorption Spectrophotometer (AAS).

### Quality control/assurance and statistical treatment of data

Analytical grade chemicals were used to prepare standard solutions and reagents. All glassware and plastics were washed with deionised water, soaked (1 + 1)HNO<sub>3</sub> overnight and finally rinsed with deionised water. Procedural blank samples were subjected to similar treatments using the same amounts of reagents. In all cases, measurements were performed in triplicate. Analysis of variance (ANOVA) was used to test differences for all investigated variables during the experiment between treatments and controls at 5% probability level ( $p \leq 0.05$ ) by means of SPSS 19.0 (IBM statistics) package.

## Results and Discussion

### Physicochemical properties of the study soil and manures

Table 1 presents some physicochemical attributes of the parent soil; manure used for the study and the metal-stressed and manure-amended composite soil collected. The parent soil and raw poultry manure are slightly acidic pH (6.21 and 6.49), therefore mobility and bioavailability of metals may not be effectively enhanced (Alkorta *et al.*, 2004) as metal solubility of Cd, Cu and Pb tends to increase at lower pH and decrease at higher pH values (Rieuwerts *et al.*, 1998). The addition of manure is expected to play indifferently a major role either in bioavailability or immobilization process of the concentration of metals (Prasad *et al.*, 2003). The bulk density, 1210 kg m<sup>-3</sup> suggested a good available water capacity, soil porosity and plant nutrient availability. Texturally, the percent of sand, silt and clay in the soil were 78.3, 13.7 and 8.33 %, patterning the soil as sandy loam. Soil organic matter and poultry manure values were 8.63 and 33.4%. Organic matter is a key for sorbing phase of metals and also plays a role in the retention of metals by soil solids, thus, decreasing mobility and bioavailability. Cation exchange capacity (CEC) is one of the important parameters in determining metal availability. The CEC for the soil and poultry manure were low (slightly saline) and this may enhance metal uptake. Therefore, organic amendments contained low levels of Cd, Cu and Pb which is suitable for use as ameliorants (Thomas and Dauda, 2015). Plant available Cd, Cu and Pb, in the soil were 7.20, 6.93 and 3.22 mg kg<sup>-1</sup>; while the pseudototal Cd, Cu and Pb contents, were 10.2, 8.93, and 5.22 mg kg<sup>-1</sup> (Table 1). Values obtained were lower than their corresponding upper critical levels, defined as the range of values above which toxicity is considered to be possible (Maiz *et al.*, 2000; Wuana *et al.*, 2012) indicating that the parent soil was relatively uncontaminated in terms of Cd, Cu and Pb. Pseudototal Cd, Cu and Pb in poultry were 10.6, 2.40 and 40.3 mg kg<sup>-1</sup>, respectively. Manure amendments decreased the plant available Cd, Cu and Pb in the soil indicating the ability

of the manures to bind the metals in the soil. Manure could sequester heavy metals and enhance fixation and immobilization in soil is recommended provided it has been allowed to age sufficiently. Therefore, soil pH, bulk density coupled with the texture suggests the potential leachability of soil heavy metals to lower profiles which makes it suitable for plant metal extraction (Wuana *et al.*, 2016).

**Table 1: Selected physicochemical properties of parent soil and poultry manure**

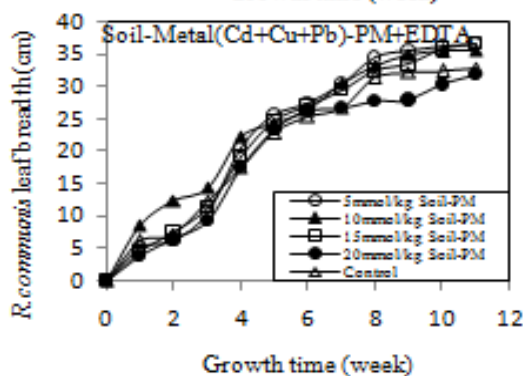
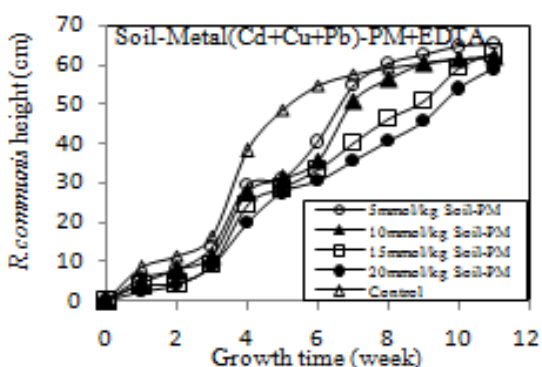
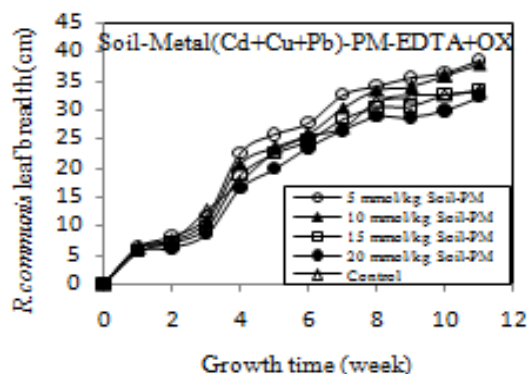
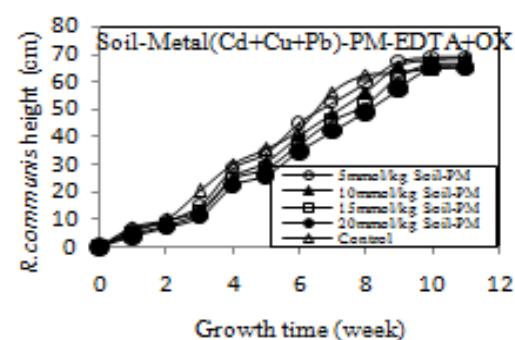
Property	Parent soil	Poultry manure
pH <sub>s</sub>	6.21 ± 0.01	6.49±0.02
Organic Matter (%)	8.63 ± 0.03	33.4±0.21
Soil Bulk Density (kg/m <sup>3</sup> )	1210 ± 1.00	
Cation exchange capacity(cmol Kg <sup>-1</sup> )	9.30 ± 0.01	
Sand (%)	78.3 ± 0.15	
Silt (%)	13.7 ± 0.03	
Clay (%)	8.33 ± 0.44	
Textural classification	Sandy loam	
<b>Available metals (mg kg<sup>-1</sup>)</b>		
Cd	7.20 ± 0.65	6.6 ±0.53
Cu	6.93 ±0.11	0.89±0.29
Pb	3.22 ±0.31	28.3±0.44
<b>Pseudo-total metal concentration (mg kg<sup>-1</sup>)</b>		
Cd	10.2±0.12	10.6±0.22
Cu	8.93 ±0.50	2.40±0.41
Pb	5.22 ±0.20	40.3±0.01

\*Mean of triplicate determinations ± standard deviation

**Growth attributes of *Ricinus communis* at increasing doses of chelants**

In this study, changes in the growth attributes (heights and leaf breadths), were investigated by pot experiments in soil stressed with Cd, Cu and Pb with increasing doses of chelants (EDTA and/or OX) with poultry manure treatments. *R. communis* plants were monitored weekly for changes in height and leaf breadth within the time stretch of 12 weeks and results illustrated in Fig. 1. At the elapse of the 90-day pot study, Soil-Metal-PM-EDTA+OX of *R. communis* height and leaf breadth were 6.23-69.0 and 4.31-38.4 cm, while that of Soil-Metal-PM-EDTA and Soil-Metal-PM+OX

were; 5.91-66.5 and 4.89-36.9 cm; 5.23-65.7 and 3.96-34.4 cm. The plants were typically greenish and luxuriant with essentially sigmoid growth profiles; i.e., plant heights and leaf breadths increased slowly in the first two weeks, followed by a sharp increase up to the fifth week and then retardation beyond this, especially in soils receiving Soil-Metal-PM-EDTA+OX treatments (Fig. 1). Chelant dose is seen to affect the growth rate and appeared to follow an approximate order: 0 mmol/kg (i.e., control) >5 (mmol/kg) >10 (mmol/kg) >15 (mmol/kg) >20 (mmol/kg). In all, the binary treatment of Soil-Metal-PM-EDTA+OX media recorded the greatest heights and leaf breadths. This could be as the result of the combine effect of chelants (EDTA and OX) where EDTA, with a large coordinating sphere complex would be quantitatively formed, leading to greater metal solubilisation and increased nutrient availability for *R. communis* (Wuana *et al.*, 2016), while the oxalic acid solubilizes metal, (Mench and Martin, 1991; Nigam *et al.*, 2001) and mobilizes mineral nutrients (Zhang *et al.*, 1989; Jones *et al.*, 1996) thereby influencing the plant growth (Reddy and Matcha, 2010; Chatzistathis and Therios, 2013). Interestingly, poultry manure enhanced the growth attributes of *R. communis*, thus, justifying its frequent land application by farmers. The fast growth rate of *R. communis* has been shown to be advantageous as a green route to carbon abatement whereby high levels of atmospheric carbon (IV) oxide are sequestered and fixed as carbon in the aerial biomass and roots (Vanaja *et al.*, 2008; Wuana *et al.*, 2016). Changes in the growth parameters with time were statistically significant (p<0.05) for all the treatments. The growth profile correlated with chelant dose and the amendment type. Growth was in the order of Soil-Metal-PM-EDTA+OX> Soil-Metal-PM-EDTA> Soil-Metal-PM-OX. Observed differences in *R. communis* heights and leaf breadths were significant (p<0.05) within and between individual potting media. However, a visual assessment of *R. communis* in the study did not show signs of phytotoxic symptoms which could alter photosynthetic order with a consequent effect on stomata or mesophyll cells in which both photochemical and biochemical reactions could be affected (Kosobrukhov *et al.*, 2004).



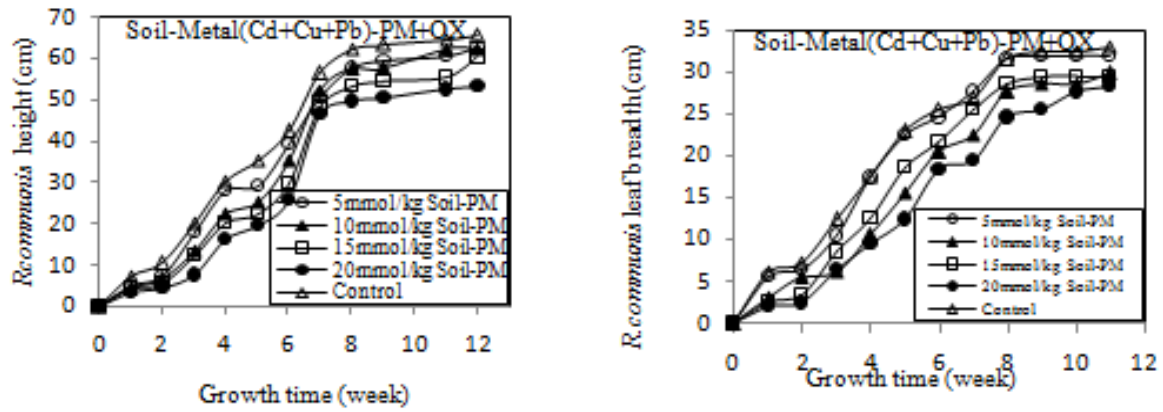


Fig. 1: Growth profile of *R. communis* in metal-poultry manure-chelant(s) amended soil

Table 2: Heavy-metal concentration in *R. communis* biomass,  $Q_p$  ( $\text{mg kg}^{-1}$ ); soil total metal,  $Q_T$  ( $\text{mg kg}^{-1}$ ); plant-available metals (calcium chloride-extracted),  $Q_e$  ( $\text{mg kg}^{-1}$ ); pH and OM (%) of poultry manure amended soil treated with different doses of chelants

Amendment Type	Dose ( $\text{mmol kg}^{-1}$ )	$Q_p$	$Q_e$	$Q_T$	pH	OM
Cd-PM EDTA+OX	5	436.0±0.41	480.1±0.31	525.3±0.33	5.80±0.12	8.71±0.14
	10	455.7±0.21	480.5±0.91	525.3±0.21	5.70±0.30	11.5±0.22
	15	477.9±0.11	481.3±0.17	518.8±0.19	5.51±0.15	13.80±0.34
	20	490.6±0.31	483.5±0.10	512.4±0.81	5.40±0.00	14.0±0.25
Cu-PM EDTA+OX	5	453.2±0.07	473.4±0.38	508.4±0.32	6.33±0.10	9.32±0.14
	10	461.7±0.54	478.1±0.11	511.7±0.11	6.38±0.34	10.7±0.30
	15	470.4±0.24	480.0±0.12	511.1±0.52	5.82±0.41	12.2±0.54
	20	475.5±0.23	480.5±0.08	502.7±0.45	5.79±0.32	13.9±0.00
Pb-PM EDTA+OX	5	452.3±0.56	471.2±0.40	522.8±0.22	6.27±0.00	8.14±0.42
	10	459.1±0.31	471.3±0.64	513.6±0.14	5.87±0.19	8.14±0.30
	15	461.9±0.41	480.5±0.66	505.9±0.36	6.01±0.20	11.2±0.11
	20	469.8±0.50	495.8±0.50	511.3±0.24	5.77±0.34	12.8±0.31
Cd-PM EDTA	5	409.1±0.14	455.2±0.54	530.7±0.18	6.59±0.31	10.2±0.21
	10	416.9±0.31	458.6±0.36	524.6±0.10	6.50±0.14	12.5±0.22
	15	427.3±0.40	457.3±0.64	516.2±0.11	6.35±0.34	12.9±0.40
	20	448.6±0.12	460.1±0.20	510.2±0.23	6.00±0.11	13.7±0.35
Cu-PM EDTA	5	312.6±0.41	461.2±0.66	529.6±0.23	6.80±0.12	10.2±0.24
	10	335.9±0.51	468.3±0.85	530.4±0.11	6.72±0.31	10.4±0.14
	15	388.8±0.20	467.6±0.01	524.2±0.87	6.50±0.20	13.7±0.40
	20	415.0±0.00	470.0±0.22	519.2±0.31	6.45±0.42	15.6±0.31
Pb-PM EDTA	5	268.6±0.60	471.2±0.64	557.7±0.21	6.82±0.45	9.88±0.14
	10	313.1±0.31	473.8±0.48	549.6±0.34	6.74±0.10	10.0±0.00
	15	374.1±0.41	475.2±0.51	539.9±0.40	6.21±0.25	13.2±0.21
	20	414.1±0.71	473.8±0.12	531.1±0.12	5.91±0.14	15.2±0.13
Cd-PM OX	5	283.5±0.04	480.1±0.34	592.9±0.54	6.99±0.41	8.4±0.33
	10	310.8±0.42	471.2±0.40	581.6±0.71	6.89±0.30	10.6±0.25
	15	356.3±0.71	473.0±0.22	580.0±0.25	6.62±0.13	12.5±0.44
	20	396.6±0.31	471.7±0.60	546.0±0.55	6.30±0.22	13.5±0.20
Cu-PM OX	5	283.5±0.26	461.2±0.24	569.7±0.18	6.72±0.10	10.2±0.42
	10	312.6±0.11	465.8±0.61	568.2±0.70	6.70±0.34	9.1±0.34
	15	345.2±0.51	466.0±0.21	566.8±0.82	6.66±0.25	11.5±0.11
	20	364.6±0.64	465.2±0.68	535.4±0.34	6.41±0.00	12.8±0.20
Pb-PM OX	5	279.9±0.30	471.2±0.42	588.6±0.41	6.24±0.12	11.2±0.14
	10	320.3±0.52	473.4±0.66	583.8±0.64	6.55±0.21	10.1±0.42
	15	361.2±0.00	471.0±0.14	577.4±0.31	5.98±0.11	12.1±0.32
	20	388.6±0.42	473.8±0.51	589.2±0.42	6.89±0.33	15.0±0.15

**Heavy metal accumulation by *Ricinus communis***

The phytoavailability of Cd, Cu and Pb to *R. communis* in response to manure and increasing levels of chelants amendment in soil was evaluated. A measure of Cd, Cu and Pb transferability to *R. communis* were calculated using the following relationship;

$$MAF = \frac{\text{Metal concentration in } R. \text{ communis tissue}}{\text{Pseudototal metal concentration in soil}} \quad (1)$$

$$RB_f = \frac{\text{Metal concentration in root of } R. \text{ communis tissue}}{\text{Pseudototal metal concentration in soil}} \quad (2)$$

$$SB_f = \frac{\text{Metal concentration in shoot of } R. \text{ communis tissue}}{\text{Pseudototal metal concentration in soil}} \quad (3)$$

$$T_f = \frac{\text{Metal concentration in shoot of } R. \text{ communis tissue}}{\text{Metal concentration in root of } R. \text{ communis tissue}} \quad (4)$$

(i) metal accumulation factor (MAF), (ii) root bioaccumulation factor (RB<sub>f</sub>), (iii) shoot bioaccumulation factor (SB<sub>f</sub>), and translocation factor (T<sub>f</sub>).

Phytoextraction can be estimated by the rate of metal accumulation and biomass production (Liang, 2009). Under Soil-Metal-PM-EDTA+OX phytoextraction, mean tissue concentrations of Cd, Cu and Pb were; 467.5, 461.8, and 464.0 mg/kg. While their respective mean pseudototal metal concentration were; 478.0, 481.4, and 479.7 mg/kg. This, however, furnished a MAF of 0.97, 0.95, and 0.96 for Cd, Cu and Pb, respectively. The uptake of Cd by the *R. communis* was higher than that of Cu and Pb. This result is consistent with the result reported by Alloway (1995) where, in his submission, averred that Cd, because of the tendency for being more mobile in the soil, is more available for plants uptake than other metals, including Pb. The concentration cadmium uptake is likely mediated through transporters or channels for other divalent ions as reported by (Cosio *et al.*, 2004). The heavy metal cadmium was depleted from the contaminated soil, suggesting absorption of cadmium metal by plant.

Root mean metal concentration for Soil-Metal-PM-EDTA+OX amended by Cd, Cu and Pb were 326.1, 321.5 and Pb 318.5 mg/kg and were greater than their corresponding mean shoot concentrations 136.8, 134.1, and 132.2 mg/kg. This however, furnished a RB<sub>f</sub>, SB<sub>f</sub> and T<sub>f</sub> of (0.68, 0.29 and 0.42); (0.67, 0.28 and 0.42); and (0.66, 0.27 and 0.41) for Cd, Cu and Pb respectively. Metal concentrations as well as RB<sub>f</sub>, SB<sub>f</sub> and T<sub>f</sub> were observed to significantly (p<0.05) increase with increasing chelant doses.

Angelova *et al.*, (2016) have reported SB<sub>f</sub> and T<sub>f</sub> values >1 for Cd and Pb in *R. communis* under natural phytoextraction. SB<sub>f</sub> and T<sub>f</sub> values of 1.03 and 1.73 have also been reported for Pb in *R. communis* under EDTA treatment or citric acid at 5.10 mmol/kg (Miniño *et al.*, 2014). The bioaccumulation of Cd, Cu and Pb were different among treatments. The metal contents in the plant tissues varied. Reason envisioned might be a function of the physiological factor of the plant, metal level uptake from solution and the xylem translocation from root to the aerial. Observed differences between the potting media were, however, not significant (p>0.05) for soil-metal-EDTA, soil-metal-OX scenarios. However, the metal concentrations in the tissue of plant appeared to follow an approximate order: EDTA+OX>EDTA>OX.

The concentrations (mg/kg) of Cd, Cu and Pb of *R. communis* grown in an array of Soil-Metal-PM-EDTA+OX, Soil-Metal-PM-EDTA, Soil-Metal-PM-OX perspective were also observed. In the Soil-Metal-PM-EDTA+OX potting scenario, tissue metal concentration varied from; 436.0-490.6 mgCdkg<sup>-1</sup>, 453.2-475.5 mgCukg<sup>-1</sup> and 452.3-469.8 mgPbkg<sup>-1</sup>, respectively. For Soil-Metal-PM-EDTA, it varied from; 409.1-448.6 mgCdkg<sup>-1</sup>, 312.5-415.5 mgCukg<sup>-1</sup>, and 268.6-414.1 mgPbkg<sup>-1</sup>. Whereas, for Soil-Metal-PM-OX, the tissue metal concentration varied from; 283.5-396.6 mgCdkg<sup>-1</sup>, 283.5-364.6 mgCukg<sup>-1</sup>, and 279.9 – 388.6 mgPbkg<sup>-1</sup> (Table 2). *R. communis* showed distinct and higher accumulation at

different chelant doses and of the amendment types. Tissue metal concentration is seen to be higher for binary chelant treatments than their corresponding single treatments. This could be probably due to the combined effect of the chelants as EDTA has the ability to raise metal concentrations in soil solutions (do Nascimento *et al.*, 2006; Zhang *et al.*, 2014; Wuana *et al.*, 2016) and Low molecular-weight organic acids i.e. oxalic acid with its high influence in the acquisition of metals by either forming complexes with metal ions where after, are extracted or desorbed from soil components (Brown *et al.*, 1994). Tissue metal concentration of treatments with EDTA was observed to have higher metal than OX. The metal-EDTA coordination sphere being larger than that of metal-OX, EDTA is hexadentate; while oxalate is bidentate (at the operating soil pH>5). For the divalent Cd, Cu, and, Pb, ions, the stoichiometric ratio of metal:EDTA in the normal octahedral complexes is always 1:1; whereas this ratio is 1:3 for the metal:oxalate complexes. Therefore, in the presence of fixed pseudototal concentration of a given metal and chelant (single or binary) dose in soil, more of the metal:EDTA complex would be quantitatively formed, leading to greater metal solubilisation and enhanced absorption by *R. communis* roots than the metal:oxalate would do (Wuana *et al.*, 2016).

Higher concentration may lead to an increase in the contents of heavy metals in tissue compartments of plants but at the same time, it could damage the growth of plants, and consequently lead to decreased accumulation of metals (Zhixin *et al.*, 2009). The criteria used for hyperaccumulation varies according the metal, ranging from 100 mgkg<sup>-1</sup> dry mass for Cd to 1000 mgkg<sup>-1</sup> for Cu, Co, Cr and Pb. These values exhibit a shoot-to-soil ratio of metal concentration and the factor for bioaccumulation is higher than 1 (Baker *et al.*, 1994). The accumulation of the selected metals varied by the nature of chelant and dosage used. Concentrations were enhanced in an approximate order: EDTA+OA>EDTA>OX. *R. communis* is a perennial plant, it is expected that the plant will be able to extract and accumulate higher amounts of the metals if allowed to stay longer in the soil (do Nascimento *et al.*, 2006). This however, indicate that *R. communis* is capable of removing the metals from the soil matrix and capable of translocating them from root to shoots.

**Conclusion**

The findings of this study have shown that the result for the physicochemical analysis of the parent soil were suitable for experimental ground. Growth of *R. communis* in the metals poultry and chelant study did not show phytotoxic symptoms and were essentially sigmoid. Poultry manure however enhanced the growth attributes. Tissue (root and shoot) of heavy metal concentrations (mg/kg) increased significantly with increased chelant doses and chelant amendment type. Altogether, the mixed chelant treatments showed enhanced metal accumulation in *R. communis* tissue to a greater extent than the single chelant scenarios, the single scenarios, EDTA showed more than oxalic acid. This suggests that the mixed chelants could be considered as alternative treatments for enhanced phytoextraction. In light of the above, the results provided ample evidence of *R. communis* as hyperaccumulator and could be very effective in the uptake of metals especially, in the presence of poultry manure and chelants.

**Acknowledgment**

We appreciate the assistance received from the staff of Chemistry laboratory, University of Agriculture, Makurdi during my laboratory work, especially Mr. Emmanuel Ukpoko for their valuable advice and technical support that has contributed to make this study a reality. Also is the staff of

Golden year limited, Port-Harcourt for carrying out Atomic Absorption Spectrometry (AAS) analysis.

#### Conflict of Interest

Authors declare that there is no conflict of interest in this study.

#### References

- Adhikari T & Kumar A 2012. Phytoaccumulation and tolerance of *Ricinus communis* L. to nickel. *International Journal of Phytoremediation* 14: 481–492.
- Ahmadpour P, Ahmadpour F, Mahmud TMM, Arifin Abdu, Soleimani M & HosseiniTayefeh F 2005. Phytoremediation of heavy metals: A green technology. *Afr. J. Biotech.*, 11(76): 14036-14043.
- Alkorta I, Hernández-Allica J, Becerril JM, Amezcaga I, Albizu I & Garbisu C 2004. Recent findings on the phytoremediation of soils contaminated with environmentally toxic heavy metals and metalloids such as zinc, cadmium, lead, and arsenic. *Revised Environ. Sci. Biotech.*, 14: 71–90.
- Alloway BJ 1995. Soil processes and the behaviour of heavy metals, pp. 11-37. In: Heavy metals in soils. Alloway B (Eds). Chapman and Hall, New York.
- Angelova V, Perifanova-Nemska M & Ivanov K 2016. Potential of castor bean (*Ricinus communis* L.) for phytoremediation of soils contaminated with heavy metals. *Int. J. Envir. Ecol. Engr.*, 3(5): 5-10.
- Baker AJM, Reeves RD & Hajar ASM 1994. Heavy metal accumulation and tolerance in British populations of the Metallophyte *ThlaspiCearu lescens* J & C Presl (*Brassicaceae*). *New Phytologist*, 127: 61 – 68.
- Blaylock MJ & Huang JW 2000. Phytoextraction of metals. In: Raskin I & Ensley BD (Eds.), phytoremediation of toxic metals: using plants to clean up the environment.– John Wiley and Sons, New York, pp. 53–70.
- Brown SL, Chaney RL, Angle JS & Baker A 1995. Chemical extractions for the assessment of heavy metal bindings in soils. *Envtal. Sci. and Techn.*, 29: 1581–1585.
- Bru¨mmer G, Gerth J & Herms U 1986. Heavy metal species, mobility and availability in soils. *Pflanzenernaehr Bodenkd.*, 147: 382-389.
- Chatzistathis T & Therios I 2013. How soil nutrient availability influences plant biomass and how biomass stimulation alleviates heavy metal toxicity in soils: The cases of nutrient use efficient genotypes and phytoremediators, respectively. Eds., Miodrag, Darko, Matovic, Biomass Now-Cultivation and Utilization,
- Cosio C, Martinoia E, Kellar C, 2004. Hyper accumulation of cadmium and zinc in *Thlaspi caerulescens* and *Arabidopsis halleri* at the leaf cellular level. *Plant Physiology*, 134: 716 – 725.
- Dembitsky V 2003. Natural occurrence of arsenic compounds in plants, lichens, fungi, algal species, and microorganisms. *Plant Science*, 165: 1177-1192.
- doNascimento CWA, Amarasiriwardena D & Xing BS 2006. Comparison of natural organic acids and synthetic chelates at enhancing phytoextraction of metals from a multi-metal contaminated soil. *Environmental Pollution*, 140: 114-123.
- Garbisu C & Alkorta I 2001. Phytoextraction: A cost-effective plant-based technology for the removal of metals from the environment. *Bioresource Technology*, 77: 229-236.
- Hornburg V & Bru¨mmer G 1993. Heavy metals in soils: Experiments on heavy metal mobility. (In German, with English abstract.) *Pflanzenernaehr. Bodenkd.*, 156: 467–477.
- Huang H, Yu N, Wang L, Gupta DK, He Z, Wang, K, Zhu Z, Yan Y, Li T & Yang X 2011. The phytoremediation potential of bioenergy crop *Ricinus communis* for DDTs and cadmium co-contaminated soil. *Bioresource Technology*, 102(23): 11034 – 11038.
- Jones DL, Darrah PR & Kochian VL 1996. Critical evaluation of organic acid mediated iron dissolution in the rhizosphere and its potential role in root iron uptake. *Plant Soil*, 180: 57–66.
- Kosobrukhov A, Knyazeva I & Mudrik V 2004. Responses to increase content of Lead in Soil: Growth and photosynthesis. *Plant Growth Regulation*, 42: 145–151.
- Liang HM, Lin TH, Chiou JM & Yeh KC 2009. Model evaluation of the phytoextraction potential of heavy metal hyperaccumulators and non-hyperaccumulators. *Environmental Pollution*, 157: 1945-1952.
- Maiz I, Arambarri I, Garcia R & Millán E 2000. Evaluation of heavy metal availability in polluted soils by two sequential extraction procedures using factor analysis. *Environmental Pollution*, 110: 3 – 9.
- McGrath SP 1998. Phytoextraction for soil remediation. In: Brooks RR (ed.) Plants that hyperaccumulate Heavy Metals: Their Role in Phytoremediation, Microbiology, Archaeology, Mineral Exploration and Phytomining. New York: CAB International, pp. 261-287.
- Mench M & Martin E 1991. Mobilization of cadmium and other heavy metals from soils by root exudates of *Zea mays* L, *Nicotiana tabacum*-L and *Nicotiana rustica* L. *Plant Soil*, 132: 187-196.
- Miniño H, Rendina A, Barros MJ, Bursztyn A, de los Ríos A, Wassner D & de Iorio AF 2014. Use of organic ligands in lead phytoextraction by castor bean (*Ricinus communis* L.). *Augmdomus*, 6: 66-80.
- Nigam R, Srivastava S, Prakash S, Srivastava MM 2001. Cadmium mobilization and plant availability – The impact of organic acids commonly exuded from roots. *Plant Soil*, 230: 107–113.
- Padmavathamma PK & Loretta YL 2007. Phytoremediation technology: Hyper-accumulation metals in plants. *Water Air Soil Pollution*, 184: 105–126.
- Pedron F & Petruzzelli G 2011. Green remediation strategies to improve the quality of contaminated soils. *Chemical Ecology*, 27: 89-95.
- Prasad MNV & Freitas H 2003. Metal hyperaccumulation in plants biodiversity prospecting for phytoremediation technology. *Electronic Journal of Biotechnology*, 6: 275-321.
- Rajkumar M & Freitas H 2008. Influence of metal resistant-plant growth promoting bacteria on the growth of *Ricinus communis* in soil contaminated with heavy metals. *Chemosphere*, 71(5): 834-842.
- Reddy KR & Matcha SK 2010. Quantifying nitrogen effects on castor bean (*Ricinus communis* L.) development, growth and photosynthesis. *Industrial Crops Production*, 31(1): 185-191.
- Revathi K, Haribabu TE & Sudha PN 2010. Phytoremediation of chromium contaminated soil using sorghum plant. *Int. J. Environ. Sci.*, 2: 417-428.
- Rieuwerts JS, Thornton I, Farago ME & Ashmore MR 1998. Factors influencing metal bioavailability in soils: Preliminary investigations for the development of critical loads approach for metals. *Chemical Speciation Bioavailability*, 10: 61-75.
- RuwanthiWettasinghe MS 2012. Development of Castor (*Ricinus communis*) Var. Brigham with Ultra Low Ricin Content by Analyzing Soluble Seed Proteins. A Dissertation In Plant and Soil Science Submitted to the Graduate Faculty of Texas Tech University in Partial

- Fulfillment of the Requirements for the Degree of Doctor of Philosophy, 57pp.
- Salt DE, Blaylock M, Kumar PBAN, Dushenkov V, Ensley BD, Chet I & Raskin I 1995. Phytoremediation: a novel strategy for the removal of toxic metals from the environment using plants. *Biotechnology*, 13: 468-475.
- Sha'Ato R, Ojanuga AG & Ajayi SO 2002. Aluminum and Fe sesquioxides dynamism in representative profiles in the Lower Benue Valley, Nigeria. *J. Agric., Sci. and Techn.*, 12: 21-31.
- Shi GR & Cai QS 2009. Cadmium tolerance and accumulation in eight potential energy crops. *Biotechnology Advances*, 27(5): 555-561.
- Thomas EY & Dauda SO 2015. Comparative effects of compost and poultry manure on bioavailability of Pb and Cu and their uptake by maize (*Zea mays* L.). *New York Science Journal*, 8(7).
- Vanaja M, Jyothi M, Ratnakumar P, Vagheera P, Reddy PR, Lakshmi NJ, Yadav SK, Maheshwari M & Venkateswarlu B 2008. Growth and yield responses of castor bean (*Ricinus communis* L.) to two enhanced CO<sub>2</sub> levels. *Plant Soil Environment*, 54(1): 38-46.
- Wang H, Shan X, Wen B, Zhang S & Wang Z 2004. Responses of antioxidative enzyme to accumulation of copper in a copper hyperaccumulator of *Compositella communis*. *Archives of Environmental Contamination and Toxicology*, 47: 185-192.
- Wuana RA, Eneji IS & Naku JU 2016. Single and mixed chelants-assisted phytoextraction of heavy metals in municipal waste dump soil by castor. *Advances in Environmental Research*, 5(1): 19-35.
- Wuana RA, Adie PA & Asegh IN 2012. Seasonal variation in bioavailability of some toxic metals in waste dump soils of Makurdi, north-central Nigeria. *J. Biodiversity Environ. Sci.*, 2(11):7-17.
- Wuana RA & Mbasugh PA 2013. Response of roselle (*Hibiscus sabdariffa*) to heavy metals contamination in soils with different organic fertilizations. *Chemical Ecology*, 29(5): 437-447.
- Wuana RA & Okieimen FE 2011. Heavy Metals in Contaminated Soils: A review of sources, chemistry, risks and best available strategies for remediation. *ISRN Ecology*, 2011: 1 – 20.
- Zhang F, Römheld V & Marschner H 1989. Effect of zinc deficiency in wheat on the release of zinc and iron mobilizing root exudates. *Z. Pflanzenernaehr. Bodenkd.*, 152: 205-210.
- Zhang H, Guo Q, Yang J, Chen T, Zhu G, Peters M, Wei R, Tian L, Wang C, Tan D, Ma J, Wang G & Wan Y 2014. Cadmium accumulation and tolerance of two castor cultivars in relation to antioxidant systems. *J. Envir. Sci.*, 26(10).
- Zhixin Niu, Lina Sun & Tieheng Sun 2009. Response of root and aerial biomass to phytoextraction of Cd and Pb by sunflower, castor bean, alfalfa and mustard. *Advances in Environmental Biology*, 3(3): 255-262.